Charcoal combined with silver for the treatment of chronic wounds

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**KEY WORDS**
- Actisorb™ dressing with silver
- Bacteria
- Chronic wounds
- Activated charcoal

**Actisorb™** dressing with silver (Systagenix Wound Management) consists principally of activated carbon impregnated with metallic silver. The carbonised fabric is enclosed in a sleeve of spun-bonded non-woven nylon, sealed along all four edges, to facilitate handling and reduce particle and fibre loss. When applied to a wound the dressing adsorbs locally released toxins and the products of wound degradation. Bacteria present in wound exudate are also attracted to the surface of the dressing where they are killed by the antimicrobial activity of the silver, which is active against a wide range of pathogenic organisms. This dressing has been used clinically for many years and, in the author’s opinion, was one of the first dressings to efficiently and favourably modulate the microenvironment of chronic wounds. The Actisorb™ dressing is marketed as ACTISORB™ Silver 220® in the United Kingdom and ACTISORB™ Ag+® in France.

Recent accumulation of fundamental data has largely clarified our understanding of the basic mechanisms that impair wound healing (Zamboni et al, 2008; Panuncialman and Falanga, 2009; Martin et al, 2009). Based on these new findings, wound management has substantially evolved. However, when reviewing these changes, it appears that the mode of action of Actisorb dressing with silver is still of clinical interest and these fundamental advances may open new ways to understand the mode of action of this dressing.

**Wound microbial flora, cytokines, inflammation and delayed healing**

Chronic wounds are contaminated by microorganisms and their bacterial ecosystem is complex (Bowler et al, 2001; Hill et al, 2003; Davies et al, 2004). For venous leg ulcers (VLU), a polymicrobial ecosystem is the rule rather than the exception (Tomijanovíc-Veselski et al, 2003; Gjødsbøl et al, 2006; Lim et al, 2006), and anaerobes may represent more than 30% of isolates from such wounds (Bowler and Davies, 1999; Halbert et al, 1992). The higher the extent of necrotic tissues in a wound, the higher the number of bacteria, whereas the wound is almost sterile when abundant granulation tissue is present (Sapico et al, 1986). While chronic wounds may tolerate large amounts of bacteria without apparent clinical signs of infection, this proliferation is not without consequences on the healing process through complex interactions with the local wound environment (Sibbald et al, 2003; Edwards and Harding, 2004; Jones et al, 2004; Ebright, 2005; Penhallow, 2005). For instance, in appropriately debrided (clear of necrotic tissue) neuropathic diabetic foot ulcers, Xu et al (2007) have shown with repeated biopsies that the wound closure rate was linearly and negatively correlated to bacterial burden. This has been confirmed by Davies et al (2007) in a prospective study conducted in 66 patients treated for VLU that were not clinically infected. The bacterial density of wound surface area measured at baseline either by swab or biopsy was a strong and independent prognostic factor of wound closure at six months. These results, among others, suggest that decreasing high bacterial burden in chronic wounds might be a favourable intervention to promote closure.

Along with metalloproteinases production enhancement, bacteria, even at low levels, have a direct effect on tissue viability via toxin secretion either directly from viable cells (exotoxins), or as a result of cell lysis (endotoxins) (Ovington, 2003). Endotoxins are known to have strong immune stimulatory and...
proinflammatory properties, even in small amounts (Fleck, 2006). At wound level, these toxins tend to cause local necrosis and will disrupt the time-regulated series of events supporting the normal healing process.

More importantly, numerous points of experimental and clinical evidence have confirmed the negative impact of bacterial biofilms (Bjarnsholt et al, 2008; Rhoads et al, 2008; Singh and Barbul, 2008; Wolcott et al, 2008). This particular bacterial mode of growth is frequent in various microorganism species and is well known for Pseudomonas aeruginosa. In wounds characterised by delayed healing and maintenance of an inappropriate inflammatory reaction, investigations have suggested that bacteria present within these wounds tend to be aggregated in microcolonies embedded in a self-produced matrix, characteristic of the biofilm mode of growth (Kirketerp-Møller et al, 2008). Additionally, there is no good correlation between bacteria detected by standard culturing methods and those detected by direct detection methods such as PNA FISH. PNA FISH uses fluorescent-labelled peptide nucleic acid (PNA) probes in a highly sensitive and specific fluorescence in situ hybridisation (FISH) assay targeting the species-specific ribosomal RNA (rRNA). This technique provides a reliable tool to study bacterial biofilm formation (Malic et al, 2009).

Therefore, toxin control is a potentially valuable adjunct to any infection control modality. A long-standing method of controlling toxins of many types is the use of high surface area adsorbents, such as activated charcoal. Data strongly suggest that activated charcoal may offer specific advantages in topical wound management through its effects on bacterial toxins, especially when combined with silver; in order to decrease bacterial burden and the risk of biofilm development (Thomas and McCubbin, 2003; Castellano et al, 2007).

Activated charcoal and silver: main properties
Activated charcoal and adsorption properties
Activated charcoal (also named as activated carbon, active carbon, or black bone) is a charcoal (most frequently of vegetable origin) that has been activated for adsorption by steaming or by heating in a vacuum. Charcoal becomes activated by heating it with steam to approximately 1000°C in the absence of oxygen (Marsh and Rodríguez-Reinoso, 2006).

Activated carbon (Figure 1) has an extraordinarily large surface area and pore volume that gives it a unique adsorption capacity (Baker et al, 1992). Some have surface areas as high as 5,000 m²/g. The specific mode of action is extremely complex and has been the subject of numerous studies and debates (Bansal and Goyal, 2005; Marsh and Rodríguez-Reinoso, 2006). Activated carbon has both chemical and physical effects on substances where it is used as a treatment agent. Activity can be usually separated into adsorption, its main property, mechanical filtration, ion exchange and surface oxidation. Activated charcoal is widely used to decolourise liquids, recover solvents, and remove toxins from water and air.

The adsorption properties of activated charcoal have been used for a long time to clear fluids from bacterial endo- and exotoxins (Du et al, 1987), and these properties have an important impact in the management of chronic wounds. For instance, in experimental conditions it has been shown that activated charcoal, when bathed in a milieu enriched with Escherichia coli

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**Figure 1. Schematic presentation of adsorption properties of activated charcoal through macro- and micropores open within the carbon matrix**

Activated charcoal is characterised by a carbon matrix incorporating a huge amount of pores of various diameters and lengths. Overall, available surface for adsorption ranges from 400 to 1,200 m² per gram of charcoal and can be up to 5,000 m².
endotoxin, will remove 90–95% of this toxin (Nolan et al, 1975; Mastra et al, 1981). In in vitro and in vivo experiments using a murine model of gut-derived endotoxaemia, charcoal particles were one of the most effective materials in the prevention of endotoxaemia. Good correlation was shown between the ability of activated charcoal to bind endotoxin in vitro as compared with in vivo action (Ditter et al, 1983). Furthermore, it has been shown that activated charcoal can also adsorb bacteria, viruses, and various other biochemicals, both in vitro and in vivo (Drucker et al, 1977; Naka et al, 2001).

Additionally, ex vivo and in vitro, activated charcoal efficiently filters clear blood from inflammatory chemokines and cytokines such as interleukin-8 (IL-8) or tumour necrosis factor-alpha (TNFα) (Cole et al, 2002; Howell et al, 2006; Sandeman et al, 2008). In an ex vivo experiment evaluating a haemofiltration device combined with an active carbon cartridge, the cartridge adsorbed 90% of IL-1beta, 72% of IL-6, 100% of IL-8, and 7% of TNFα during each pass (Cole et al, 2002).

Malodour is a common problem when managing wounds, especially malignant wounds or pressure ulcers (Lee et al, 2006; McDonald and Lesage, 2006; Seaman, 2006). This problem may have a detrimental impact on patients’ body image (Young, 2005). The blood supply to parts of the tumour or of the wound is often impaired, resulting in areas of hypoxic or necrotic tissue. These areas become infected with anaerobic bacteria, which release malodorous volatile fatty acids as a metabolic by-product. Activated charcoal heavily adsorbs and neutralises these volatile molecules, thus decreasing odour as shown in a double-blind experiment (Chakravarthi et al, 2008). This will help both patients and healthcare professionals to more easily comply with wound care.

Advantages of adding silver
Silver has a broad antibacterial spectrum including all microorganisms involved in wound colonisation and wound infection (Lansdown, 2002). Due to its mechanisms of action, resistance to silver is hard to develop (Percival et al, 2005). In addition, silver possesses other properties of clinical interest in wound management. It is a strong anti-inflammatory agent and is able to inhibit matrix metalloproteinase activity (Kerihuel, 2008).

Silver incorporated in the activated charcoal matrix is primarily bound to the structure of the charcoal and not released. However, in conjunction with bacterial adsorption onto charcoal surface, silver ions in the material kill these bacteria and efficiently participate to reduce bacterial burden and biofilm growth (Muller et al, 2003).

Actisorb in the management of chronic wounds: clinical evidence
Randomised controlled trials of Actisorb without silver
Two randomised control studies were conducted with Actisorb in the treatment of chronic wounds.

Mulligan et al (1986) randomised a total of 101 subjects with ulcers to either Actisorb or to any other dressing regarded as most appropriate by investigators (control group). These various ulcers were treated for six weeks, or until healing. Main evaluation parameters included ulcer size and wound aspect (granulation tissue, exudate level, oedema, odour, pain). There were no statistically significant differences in baseline demographic data. However, the Actisorb group included ulcers of longer mean duration (117 versus 52 months), and larger mean area (30.3 cm² versus 19.7 cm²). In the Actisorb group, a statistically significant 28.7% (±3.9%) mean percentage reduction in wound area was noted compared to 1.7% (±6.8%) reduction in the control group. In addition, Actisorb was statistically superior for reduced exudate, odour and oedema levels (p=0.005).

Another randomised controlled trial was conducted in pressure ulcers (Kerihuel et al, 2003). The wounds included in the study were moderately exudative but not clinically infected. The control group was treated with a hydrocolloid dressing. Patients were followed up to healing or up to eight weeks. In the study group, Actisorb was used initially and then switched to an interface dressing (Adaptic [Johnson & Johnson]) as soon as complete debridement was achieved. Wounds were medically assessed weekly and all local care was recorded. Sixty-two patients were included in the study. Wound area decreased similarly with both dressing strategies (centralised and blind measurement of wound tracings). Time to reach debridement was, however, shorter with Actisorb (26 ±9 days versus 31 ±5 days) days). Effective debridement was defined as a wound with no or very weak exudation, less than 25% of its surface still covered by slough and no more black necrotic tissue. Achievement of debridement was to be documented by a picture. Local tolerance of applied dressings was good in both groups.

Randomised controlled trials with Actisorb with silver
Millward (1991) compared Actisorb dressing with silver with two control dressings (a chlorhexidine paraffin gauze and a charcoal dressing) on 60 patients with leg ulcers. All wounds were swabbed at weekly intervals for two weeks before the study start and for four weeks with the test dressings in use. Wounds were assessed subjectively at each dressing change according to a standard scale. The overall assessments of dressing performance supported a higher efficiency of Actisorb dressing with silver wound management in wound cleansing, malodour control and exudate reduction. All Actisorb treated wounds showed healing improvement.

Wunderlich and Orfanos (1991) treated 40 patients with chronic VLU. They were randomised to either conventional (zinc paste bandages) or test (Actisorb dressing with silver) therapy. During the six-week treatment period, wound parameters were measured (ulcer size, odour, necrotic tissue, epithelialisation, erythema, oedema). Nineteen patients completed the treatment period in each group. Actisorb dressing with silver was statistically superior to the control (p<0.05) in terms of wound area reduction with six ulcers (32%) healed, compared with two (10%) in the control group.
In another randomised trial conducted in venous leg ulcers (VLU) (Keniuel and Dujardin-Detrez, 2003), controlled groups were treated with a hydrocolloid dressing. Patients were followed up to healing or up to eight weeks. In the study group, Actisorb dressing with silver was used initially and then switched to an interface dressing (Adaptic) as soon as complete debridement was achieved. Wounds were medically assessed weekly and all local care was recorded. Sixty patients were enrolled and all received compression bandaging. At one week visit, median regression of wound area was 15% in the Actisorb dressing with silver group and 3% in the control group (p=0.07); A 50% or more area reduction was observed in 53.3% of the patients treated with Actisorb, as compared with 46.7% in the hydrocolloid group.

Finally, Verdu Soriano et al (2004) confirmed that Actisorb dressing with silver reduced bacterial burden. In this trial, the effect of Actisorb dressing with silver in reducing the level of bacteria in chronic wounds with no clinical signs of local infection was compared with cleansing and debridement (control group). Patients were randomly assigned to the intervention or control group and followed over two weeks. Surface smear (spatula) and percutaneous aspiration were performed at baseline and after 15 days. Sixty-seven lesions were included in the intervention group and 58 in the control group. At baseline in the intervention group, 71.6% of the wounds were colonised (≤10 cfu colony forming unit [CFU]), 7.5% had a high level of bacteria (≥10⁵ CFU) and 20.9% were bacteriologically considered infected (≥10⁸ CFU). At baseline in the control group, 65.5% of the wounds were contaminated, 13.8% colonised or had a high level of bacteria, and 20.7% were infected. After two weeks, 85.1% (57/67) of the contaminated/colonised wounds in the intervention group had a reduction in the number of bacteria compared with 62.1% (36/58) in the control group (p=0.003). For infected wounds at baseline, the bacterial level was still ≥10⁵ CFU at two weeks in 7.1% (1/14) and 75.0% (9/12) in the Actisorb dressing with silver and control groups respectively (p=0.001).

**Other clinical evidence**

Other data from non-controlled studies and large population surveys or case reports support the clinical benefit of using Actisorb dressing with silver in various clinical situations (White, 2001; Keniuel and Dujardin-Detrez, 2003).

Furthermore, some evidence may support extension of the use of Actisorb dressing with silver to other types of wounds. As documented by Leak et al (2002), any breach in the integrity of the skin (e.g. incisions, intravascular catheters) can act as a portal for the ingress of microorganisms and thereby predispose the patient to infection. The infection, if unchecked, can put the patient at risk of bacteraemia. In cases of percutaneous enterostomal gastrostomy (PEG) sites, implementation of a protocol for skin care around PEG sites, including the use of Actisorb dressing with silver, indicates that patient comfort can be improved, hypergranulation reduced, and mexitilline-resistant Staphylococcus aureus colonisation and infection eradicated (Leak et al, 2002).

**Conclusion**

Actisorb dressing with silver consits principally of activated carbon impregnated with metallic silver. This dressing combines the adsorption capacity of activated charcoal to the large-spectrum antibacterial properties of silver. In the author’s practical experience, Actisorb actively modulates the impaired wound healing process of chronic wounds. Its main property is its potential to adsorb and neutralise the bacterial toxins locally released and to reduce the inappropriate high level of cytokines in the wound exudate of chronic wounds. In addition, incorporating silver actively participates in decreasing bacterial bioburden and biofilm growth, a major factor of chronicity.

Overall, the various studies discussed above support that Actisorb dressing with silver is a therapeutic alternative when treating chronic wounds that are stuck in an inflammatory stage. It may be used in the treatment of most types of chronic wounds but seems particularly useful for the management of strongly colonised wounds, and even of heavily exuding ulcers if combined with a high-absorbing secondary dressing such as a foam dressing.  

**References**


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**Product REVIEW**

**Key points**

- Activated charcoal has huge adsorption capacities for wound debris, bacterial species and bacterial toxins as well as for volatile amines.

- By combining silver to charcoal, this dressing will decrease local bacterial burden.

- Actisorb Silver is an efficient tool to help manage debridement of chronic wounds.

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